



## Original Article

# EVALUATION OF ANTIHYPERLIPIDEMIC, ANTI-INFLAMMATORY, AND ANALGESIC ACTIVITIES OF *EURYCOMA LONGIFOLIA* IN ANIMAL MODELS

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## ABSTRACT

**Objective:** To investigate the anti-hyperlipidemic, anti-inflammatory and analgesic properties of *E. longifolia* root extract in animal models.

**Methods:** In this study, glucose-fructose enriched diet-induced hyperlipidemia, carrageenan-induced paw edema and acetic acid-induced writhing were used to evaluate the anti-hypertriglyceridemia, anti-inflammatory and analgesic activities, respectively. At the end of the experiment of glucose-fructose enriched diet-induced hyperlipidemia, blood samples were collected and estimation of blood lipids were carried out. Edema thickness was measured using digital caliper at 0, 15, 30, 45, 60, 90, 120, 150, 180, 210, 240, 270, 300, 330, and 360 min after carrageenan injection. The number of abdominal writhing for each mouse was observed and counted during a period of 1 h post injection of acetic acid.

**Results:** *E. longifolia* root extract demonstrated a significant reduction of triglyceride levels ( $p < 0.05$ ) compared with the control group in glucose-fructose enriched diet in rats. In anti-inflammatory test, the extract significantly inhibited the carrageenan induced paw edema formation ( $p < 0.05$ ). The extract also significantly decreased the number of writhing in acetic acid-induced mice ( $p < 0.05$ ).

**Conclusion:** *E. longifolia* root extract shown a significant anti-hypertriglyceridemia, anti-inflammatory and analgesic activities. Further studies are needed to determine mechanisms for its activities of *E. longifolia* root extract.

**Keywords:** *Eurycoma longifolia*, Antihypertriglyceridemia, Anti-inflammatory, Analgesic

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## INTRODUCTION

Obesity and the metabolic syndrome continue to plague the world at an alarming rate. Those will cause both substantial socio-economic and physical burden in society. WHO reported that in 2014, over 1.9 billion adults were estimated to be overweight and more than 600 million obese [1].

Recently people have been using natural products and plant to treat a wide variety of clinical disease. One of the most popular traditional plants *Eurycoma longifolia* Jack (Simaroubaceae) is a well-known folklore herbal medication in Southeast Asia. Wide spectrum of pharmacological activities of *E. longifolia* has been reported [2-5]. Despite wide range of traditional uses known, it is most solely for its aphrodisiac and anti-malarial properties [5-7]. Previous research shows that extracts from the roots of *E. longifolia* suppressed intracellular lipid accumulation in 3T3-L1 adipocytes, a treatment target for an anti-obesity agent [8]. Based on the previous study and traditional use, the present investigation was carried out to evaluate the antihyperlipidemic, anti-inflammatory and analgesic activities of *E. longifolia* root extract in animal models.

## MATERIALS AND METHODS

### Plant material and chemicals

*E. longifolia* was supplied by Merapi Farma Herbal Co. (Batch No. SL.1A.2015. PB; Yogyakarta, Indonesia) and were collected from Kalimantan Island Indonesia. The voucher specimen was deposited at the Laboratory of Pharmaceutical Biology, Faculty of Pharmacy Sanata Dharma University in Yogyakarta Indonesia. The following chemicals were used: sodium carboxy methyl cellulose/CMC-Na (Brataco Chemika, Indonesia); glucose, fructose, methanol and acetic acid (E. Merck, Darmstadt Germany; and carrageenan (Sigma Chemical Company). Diagnostic kit for the estimation of Cholesterol, high-density lipoprotein cholesterol (HDL-c), low-density lipoprotein cholesterol (LDL-c), triglyceride (TG) kits were purchased from Roche Diagnostics GmbH, Mannheim, Germany. All other chemical were of analytical grade and were purchased from E. Merck,

Darmstadt, Germany. Instruments used in this study included Cobas C501 (Roche Diagnostics) for serum chemistry analysis.

### Preparation of *E. longifolia* root extract

The root of *E. longifolia* were powdered and extracted with 95% (v/v) methanol for 48 h at room temperature. The methanol extract was then distilled, evaporated to obtain semisolid *E. longifolia* root extract (EL) (yield 9.32%) and re-dissolved in CMC-Na (1% w/v).

### Test animals and housing

28 adult male Wistar rats (150-250 g), and 25 adult male and 25 female Swiss mice (20-30 g) were obtained from the Imono Laboratory, Sanata Dharma University, Indonesia. The animals were maintained under standard laboratory condition. They were housed in standard cages (five animals per cage) at temperature  $22 \pm 2^\circ\text{C}$  and 12:12h light dark cycle. The animals were provided with pelleted diet as normal diet or glucose-fructose enriched diet (GFED) and water ad libitum. All procedures described were reviewed and approved with approval number KE/KF/337/EC by Medical and Health Research Ethics Committee Faculty of Medicine Gadjah Mada University Yogyakarta Indonesia.

### Antihyperlipidemic study

Healthy male rats were fasted overnight and randomly divided into four groups each containing 7 animals. The control group was fed normal diet until the end of treatment. The remaining groups were fed GFED for 42 d [9]. Following confirmation of GFED-induced hypertriglyceridemia, the three groups were then divided into group I (EL 75 mg/kg b/w), group II (EL 150 mg/kg b/w) [10], and group GFED-control (continued with vehicle). All treatments were continued for 5 d following oral administration. At the end of treatment, rats in all groups were anaesthetized with ketamine. The blood was collected from all groups of rats by retro-orbital sampling for serum chemistry analysis. Serum lipid profile tests were measured using commercial kits (Roche Diagnostics).

### Anti-inflammatory study

Male Swiss mice were divided into five groups randomly (negative control, positive control and dose groups), consisting of 5 mice in each group. Acute edema was induced by the injection of carrageenan 1% (prepared in normal saline) into the sub-plantar region of hind-paw of mice [11]. Then, each group was treated orally with 1% CMC-Na (negative control), 4.48 mg/kg BW of diclofenac sodium (positive control), and treatment doses of 105, 210 and 420 mg/kg BW EL [10]. Edema thickness was measured using digital caliper at 0, 15, 30, 45, 60, 90, 120, 150, 180, 210, 240, 270, 300, 330, and 360 min after carrageenan injection [12, 13]. The calculation of the edema volume was conducted using formula area under curve (AUC) and percentage of inhibition of inflammation [13-15].

Area under curve (AUC) was calculated for each minute within 0-6 h using trapezoid method formula, as below:

$$AUC_{t_n-1} = \frac{t_n}{2} \frac{T_{t_n-1} + T_{t_n}}{(t_n - t_{n-1})} \quad [16]$$

$T_{t_n-1}$ : Average edema volume on  $t_{n-1}$

$T_{t_n}$ : Average edema volume on  $t_n$

### Analgesic study

The analgesic activity of EL was tested using acetic acid-induced writhing method [11, 14, 15, 17]. Female mice Swiss were divided randomly into five groups (n=5). Group I as negative control

received 1% CMC-Na, group II as positive control received 91 mg/kg BW of aspirin, group III-V received 105, 210 and 420 mg/kg BW of EL respectively. After 5 min of treatment, all animals were administered 1% with 50 mg/kg BW of acetic acid intraperitoneally. The total number of writhes was recorded at intervals of 5 min for a period of 1 h. The calculation of percentage of inhibition using the following ratio: (control mean-treated mean)/control mean x 100 [14, 15].

### Statistical analysis

Results are expressed as mean±standard deviation (SD). Data were analyzed using one-way analysis of variance followed by *post-hoc* Tukey HSD tests using SPSS 22. A *p*-value<0.05 was considered statistically significant. Statistical differences were determined using the Student's *t*-test, with *p*-values being indicated for each fig.

### RESULTS

#### Effect of *E. longifolia* root extract in hyperlipidemia-induced rats

The administration of GFED significantly increased (*p*<0.05) TG levels up to 186.1% and decreased high-density lipoprotein (HDL) levels by 43.0% (table 1). GFED did not increase serum total cholesterol or low-density lipoprotein (LDL) levels.

Daily administration of both doses of EL for 5 d led to a significant reduction (*p*<0.05) in TG compared with the GFED group. There was no significant difference in HDL levels among all doses of EL and those of the GFED group.

**Table 1: Effect of *E. longifolia* root extract on lipid parameters in rats feeds with glucose-fructose enrich diet (GFED)**

Treatment	Cholesterol (mmol/l)	Triglyceride (mmol/l)	HDL-c (mmol/l)	LDL-c (mmol/l)
Normal diet	1.74±0.16	1.08±0.09 <sup>b</sup>	1.65±0.08 <sup>b</sup>	0.21±0.05
GFED	1.86±0.15	3.08±0.21 <sup>a</sup>	0.94±0.07 <sup>a</sup>	0.24±0.04
GFED+EL 75 mg/kgBW	1.69±0.26	2.69±0.34 <sup>a,b</sup>	0.92±0.07 <sup>a</sup>	0.25±0.06
GFED+EL 150 mg/kgBW	1.88±0.17	0.43±0.05 <sup>a,b</sup>	0.70±0.08 <sup>a,b</sup>	0.37±0.07 <sup>a,b</sup>

Values are expressed as mean±SD of seven animals in each group; a: *p*<0.05 vs normal diet; b: *p*<0.05 vs GFED

#### Effect of *E. longifolia* root extract on carrageenan-induced mice

The anti-inflammatory activity of *E. longifolia* root extract against carrageenan induced paw edema showed that the extracts exhibit

significantly (*p*<0.05) and dose-dependently reduced the paw edema swelling (table 2). The percentage inhibition in the paw edema in mice treated with *E. longifolia* root extract was found to be 15.7; 22.0 and 26.8% at the dose of 105; 210 and 420 mg/kg BW respectively.

**Table 2: Effect of *E. longifolia* root extract on carrageenan-induced mice**

Treatment	AUC	% inhibition
Negative control (CMC Na)	426.7 ± 18.3 <sup>b</sup>	0.0
Positive control (diclofenac sodium 4.48 mg/kg BW)	255.9 ± 16.8 <sup>a</sup>	40.0
EL 105 mg/kg BW	359.6 ± 5.8 <sup>a,b</sup>	15.7
EL 210 mg/kg BW	333.0 ± 5.6 <sup>a,b</sup>	22.0
EL 420 mg/kg BW	312.4 ± 4.0 <sup>a,b</sup>	26.8

Values are expressed as mean±SD of five animals in each group; a: *p*<0.05 vs negative control; b: *p*<0.05 vs positive control

#### Effect of *E. longifolia* root extract on acetic acid-induced writhing in mice

Table 3 showed the effect of *E. longifolia* root extract on acetic acid-induced writhing in mice. All of the doses of *E. longifolia* root extract

produced significant (*p*<0.05) reduction of writhing by the acetic acid in a dose dependent manner.

The percent inhibition of 105, 210 and 420 mg/kg BW was 25.4; 57.3 and 59.0% for *E. longifolia* root extract respectively.

**Table 3: Effect of *E. longifolia* root extract on acetic acid induced writhing in mice**

Treatment	Number of writhing	% inhibition
Negative control (CMC Na)	57.6 ± 1.1 <sup>b</sup>	0.0
Positive control (aspirin 91 mg/kgBW)	16.2 ± 1.3 <sup>a</sup>	71.9
EL 105 mg/kg BW	43.0 ± 1.0 <sup>a,b</sup>	25.4
EL 210 mg/kg BW	24.6 ± 2.1 <sup>a,b</sup>	57.3
EL 420 mg/kg BW	23.6 ± 4.2 <sup>a,b</sup>	59.0

Values are expressed as mean±SD of five animals in each group; a: *p*<0.05 vs negative control; b: *p*<0.05 vs positive control

## DISCUSSION

Several studies have reported that a high carbohydrate diet is responsible for the development of hypertriglyceridemia in rodent animal models [9, 18-20]. Hypertriglyceridemia occurs because high fructose in the blood leads to increased *de novo* hepatic fatty acid synthesis and, subsequently, this releases a high amount of TG [21]. The *E. longifolia* root extract used in our study reduced TG levels induced by glucose-fructose enrich diet. Therefore, the *E. longifolia* root extract has a potent anti-hypertriglyceridemia activity in rats. These reductions of TG may be associated with a previous *in vitro* report in which *E. longifolia* suppressed lipid accumulation in 3T3-L1 adipocytes [8].

Carrageenan, as irritant substances, induced inflammation in biphasic event. The initial phase is associated to the release of serotonin, histamine, and bradykinin; while the late phase is attributed to the release of prostaglandin and induce of cyclooxygenase that increasing vascular permeability and the neutrophil infiltration into the inflammatory site and production of free radicals that cause edema [22]. In our results, the *E. longifolia* root extract significantly inhibited paw edema-induced carrageenan in all the dose level.

Additionally, the *E. longifolia* root extract showed significant analgesic action at all dose levels (105, 201 and 420 mg/kg BW). Analgesic effect was evaluated using acetic acid-induced writhing test in mice. Acetic acid injection has been associated with increased level of E and F prostaglandins in peritoneal fluids as well as lipooxygenase products [23]. The significant reduction of *E. longifolia* root extract might be due to the presence of analgesic principles acting with the prostaglandin pathways.

It has been reported that *E. longifolia* contains quassinoids, triterpene, biphenylneolignan and alkaloid [7, 24-26]. The presence of eurycomalactone, 14,15 $\beta$ -dihydrokleanone and 13, 21-dehydroeurycomanone in *E. longifolia* root were identified as potent NF- $\kappa$ B inhibitors [27, 28]. They act by inactivation of the NF- $\kappa$ B signaling pathway, a pro-inflammatory transcriptional factor. Therefore, their inhibition results in anti-inflammatory effect. Anti-inflammatory and analgesic effects of *E. longifolia* root extract in this study are in agreement with a previous study. Han *et al.* demonstrated that methanolic extract of *E. longifolia* root has a potential analgesic agent on both heat-induced pain and chemical induced pain in hot plate test and acetic acid-induced writhing test, respectively. Methanolic extract of *E. longifolia* root also has anti-inflammatory agent on carrageenan-induced paw edema [10].

## CONCLUSION

In conclusion, we can confirm that *E. longifolia* root extract are endowed with anti-hypertriglyceridemic, anti-inflammatory and analgesic properties that support to the traditional use of this plant. However, further study is needed to investigate the mechanisms of pharmacological effects.

## CONFLICTS OF INTERESTS

The authors declare that they have no conflicts of interest

## REFERENCES

- WHO. Obesity and Overweight; 2015. Available from: <http://www.who.int/mediacentre/factsheets/fs311/en/>. [Last accessed on 20 Oct 2016]
- Mohamed AN, Vejayan J, Yusoff MM. Review on *Eurycoma longifolia* pharmacological and phytochemical properties. J Appl Sci 2015;15:831-44.
- Al-Salahi OSA, Lam CK, Majid AMSA, Al-Suede FSRA, Saghir SAM, Abdullah WZ, *et al.* Anti-angiogenic quassinoid-rich fraction from *Eurycoma longifolia* modulates endothelial cell function. Microvasc Res 2013;90:30-9.
- Jiwajinda S, Santisopasri V, Murakami A, Sugiyama H, Gasquet M, Riad E, *et al.* *In vitro* anti-tumor promoting and anti-parasitic activities of the quassinoids from *Eurycoma longifolia*, a medicinal plant in Southeast Asia. J Ethnopharmacol 2002;82:55-8.
- Rashid M, Kumar S, Ahmad B. Medical uses of *Eurycoma longifolia* Jack: a review. Pharm Res 2009;2:70-8.
- Chan KL, Choo CY, Abdullah NR, Ismail Z. Antiplasmodial studies of *Eurycoma longifolia* jack using the lactate dehydrogenase assay of *Plasmodium falciparum*. J Ethnopharmacol 2004;92:223-7.
- Chua LS, Amaiza N, Amin M, Chun J, Neo H, Lee TH, *et al.* LC-MS/MS-based metabolites of *Eurycoma longifolia* (Tongkat Ali) in Malaysia (Perak and Pahang). J Chromatogr B: Anal Technol Biomed Life Sci 2011;879:3909-19.
- Lahrita L, Kato E, Kawabata J. Uncovering potential of Indonesian medicinal plants for glucose uptake enhancement and lipid suppression against 3T3-L1 adipocytes. J Ethnopharmacol 2015;168:229-36.
- Hendra P, Jamil OA, Maharani DA, Suhadi MA, Putri CY, Fenty, *et al.* Anti-hyperlipidemic and hepatoprotective studies on leaves of *Macaranga tanarius*. Asian J Pharm Clin Res 2017;10:1-3.
- Han YM, Woo SU, Choi MS, Park YN, Kim SH, Yim H, *et al.* Anti-inflammatory and analgesic effects of *Eurycoma longifolia* extracts. Arch Pharm Res 2016;39:421-8.
- Chamundeeswari D, Vasantha J, Gopalakrishnan S, Sukumar E. Anti-inflammatory and antinociceptive activities of *Trewia polycarpa* roots. Fitoterapia 2004;75:740-4.
- Belemkar S, Thakre SA, Pata MK. Evaluation of anti-inflammatory and analgesic activities of methanolic extract of *Adhatoda vasica* nees and *Mentha piperita* Linn. Inventi J 2013;2:1-6.
- Boakye-Gyasi E, Woode E, Ainooson GK, Obiri DD, Ansah C, Duwejua M, *et al.* Anti-inflammatory and antipyretic effects of ethanolic extract of *Palisota hirsuta* K. schum roots. Afr J Pharm Pharmacol 2008;2:191-9.
- Nguemfo EL, Dimo T, Azebaze AGB, Asongalem EA, Alaoui K, Dongmo AB, *et al.* Anti-inflammatory and anti-nociceptive activities of the stem bark extract from *allanblackia monticola* STANER L. C. (Guttiferae). J Ethnopharmacol 2007;114:417-24.
- Chaulya NC, Haldar PK, Mukherjee A. Anti-inflammatory and analgesic activity of methanol extracts of *Cyperus tegetum* Roxb. Rhizome. J PharmaSciTech 2012;1:27-9.
- Sinatra RS, Jahr JS, Watkins-Pitchford JM. editors. The Essence of Analgesia and Analgesics. London: Cambridge University Press; 2011.
- Rahayu L, Dewi RS, Ayu G. Anti-inflammation and analgesic test effect of senggani leaves (*Melastoma malabathricum* L.) infusion. Indonesian J Pharm Sci 2016;14: 93-8.
- Huang IS, Ho H, Hoffman BB, Reaven GM. Fructose-induced insulin resistance and hypertension in rats. Hypertension 1987;10:512-6.
- Padiya R, Khatua TN, Bagul PK, Kucha M, Banerjee SK. Garlic improves insulin sensitivity and associated metabolic syndromes in fructose fed rats. Nutr Metab 2011;8:53.
- Tandrasasmita OM, Wulan DD, Nailufar F, Sinambela J, Tjandrawinata RR. Glucose-lowering effect of DLBS3233 is mediated through phosphorylation of tyrosine and upregulation of PPAR $\gamma$  and GLUT4 expression. Int J Gen Med 2011;4:345-7.
- Huang D, Dhawan T, Young S, Yong WH, Boros LG, Heaney AP. Fructose impairs glucose-induced hepatic triglyceride synthesis. Lipid Health Disease 2011;10:1-10.
- Necas J, Bartosikova L. Carrageenan: a review. Vet Med 2013;58:187-205.
- Deraedt R, Jouquey S, Delevallee F, Flahaut M. Release of prostaglandins e dan f in an algogenic reaction and its inhibition. Eur J Pharmacol 1980;61:17-24.
- Ang HH, Hitotsuyanagi Y, Fukuya H, Takeya K. Quassinoids from *Eurycoma longifolia*. Phytochemistry 2002;59:833-7.
- Huyen LT, Nhiem NX, Thu VK, Tai BH, Anh HLT, Yem PH, *et al.* Quassinoids from *Eurycoma longifolia*. Vietnam J Chem 2015;53:82-5.
- Park S, Nhiem NX, Kiem PV, Minh CV, Tai BH, Kim N, *et al.* Five new quassinoids and cytotoxic constituents from the roots of *Eurycoma longifolia*. Bioorg Med Chem Lett 2014;24:3835-40.
- Hajjoui S, Chateauvieux S, Teiten MH, Orlikova B, Schumacher M, Dicato M, *et al.* Eurycomanone and eurycomanol from

*Eurycoma longifolia* jack as regulators of signaling pathways involves in proliferation, cell death and inflammation. Molecules 2014;19:14649-66.

28. Tran TVA, Malainer C, Schwaiger S, Atanasov AG, Heiss EH, Dirsch VM, *et al.* NF- $\kappa$ B inhibitors from *Eurycoma longifolia*. J Nat Prod 2014;77:483-8.

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